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**INDO AMERICAN JOURNAL OF
PHARMACEUTICAL SCIENCES**<http://doi.org/10.5281/zenodo.1160874>Available online at: <http://www.iajps.com>**Review Article****A REVIEW ON ZIKA VIRUS ASSOCIATED MICROCEPHALY****Reza Rajesh^{1*}, Sharon Liza Koshy², Renuka R³, Dr. Elesy Abraham⁴**¹Third Year Pharm D Student, Nazareth College of Pharmacy, Othera P.O, Thiruvalla, Kerala, India²Third Year Pharm D Student, Nazareth College of Pharmacy, Othera P.O, Thiruvalla, Kerala, India³Assistant Professor, Nazareth College of Pharmacy, Othera P.O, Thiruvalla, Kerala, India⁴Principal, Nazareth College of Pharmacy, Othera P.O, Thiruvalla, Kerala, India**Abstract:**

Microcephaly is a condition where a baby's head is much smaller than expected. During pregnancy, a baby's head grows as brain is enlarging. Microcephaly can occur because a baby's brain has not developed properly during pregnancy or has stopped growing after birth, which results in a smaller head size. Zika virus is a mosquito-borne flavivirus transmitted primarily by Aedes aegypti mosquitoes. These vectors also transmit dengue and chikungunya virus and are found throughout much of the Americas, including parts of the United States. Zika virus infections have been confirmed in infants with microcephaly. In the current outbreak in Brazil, a marked increase in the number of infants born with microcephaly has been reported. Studies are under way to investigate the association of Zika virus infection and microcephaly. There are currently no drugs approved for the treatment of ZIKV-infection. Clinical trials are going on for the development of Zika vaccine. The aim of drug development is primarily to reduce viral load, reduce symptoms, and protect the unborn fetus from neurological sequelae. In order to develop effective anti-ZIKV vaccines and therapeutics, improved animal models and a better understanding of immunological correlates of protection against ZIKV are required. This review will summarize what is currently known about ZIKV, the clinical manifestations and epidemiology of Zika as well as, host immune responses against ZIKV, and the current state of development of vaccines and therapeutics against ZIKV.

Key Words: zikavirus (ZIKV), microcephaly, zikavaccine, flavivirus, Aedes aegypti***Correspondence to Author :****Ms. Reza Rajesh,**

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INTRODUCTION:

Zika is a rapidly emerging public health threat. The Zika virus has spread rapidly in the Americas since its first identification in Brazil in early 2015. Adverse effects in pregnancy such as microcephaly and other serious brain anomalies and birth defects were found to be linked with prenatal Zika virus infection. Zika virus is a mosquito-borne flavivirus transmitted primarily by *Aedes aegypti* mosquitoes. [1,24]

Almost 80% of persons infected with Zika virus are asymptomatic. Most commonly found symptoms are acute onset of fever, maculopapular rash, arthralgia, or non-purulent conjunctivitis. The primary mode of ZIKV transmission is through the bite of infected mosquitoes, *Aedes aegypti* and *Aedes albopictus*. They are considered as the predominant vectors. Other than vector-borne transmission, sexual transmission also contributes to Zika virus spread. Other modes of transmission include transmission from mother to child, blood transfusion, laboratory transmission, and transmission by physical contact [9,16,18,19,21,22,28,42].

EPIDEMIOLOGY

From the reports of human cases of ZIKV infection from 1947 until the 2007 it was found that ZIKV has now spread dramatically to over 80 countries and territories with vector-borne transmission. ZIKV was first reported in Brazil in 2015 with large numbers of suspected cases and the initial observation that the number of newborn infants with microcephaly was increased in ZIKV-affected areas later that year. The studies conducted in French Polynesia in 2013-2014 revealed that the risk of microcephaly due to ZIKV infection in the first trimester of pregnancy was 0.95%. According to Zika virus outbreak which began in October, 2013, and ended in April, 2014, it was estimated that 66% of the general population were infected. Among a cohort of 35 infants with microcephaly born during August–October 2015 in eight of Brazil's 26 states and reported to the registry, the mothers of all 35 had lived in or visited Zika virus-affected areas during pregnancy, 25 infants had severe microcephaly, 17 had at least one neurologic abnormality, and among 27 infants who had neuroimaging studies, all had abnormalities [11,24,49].

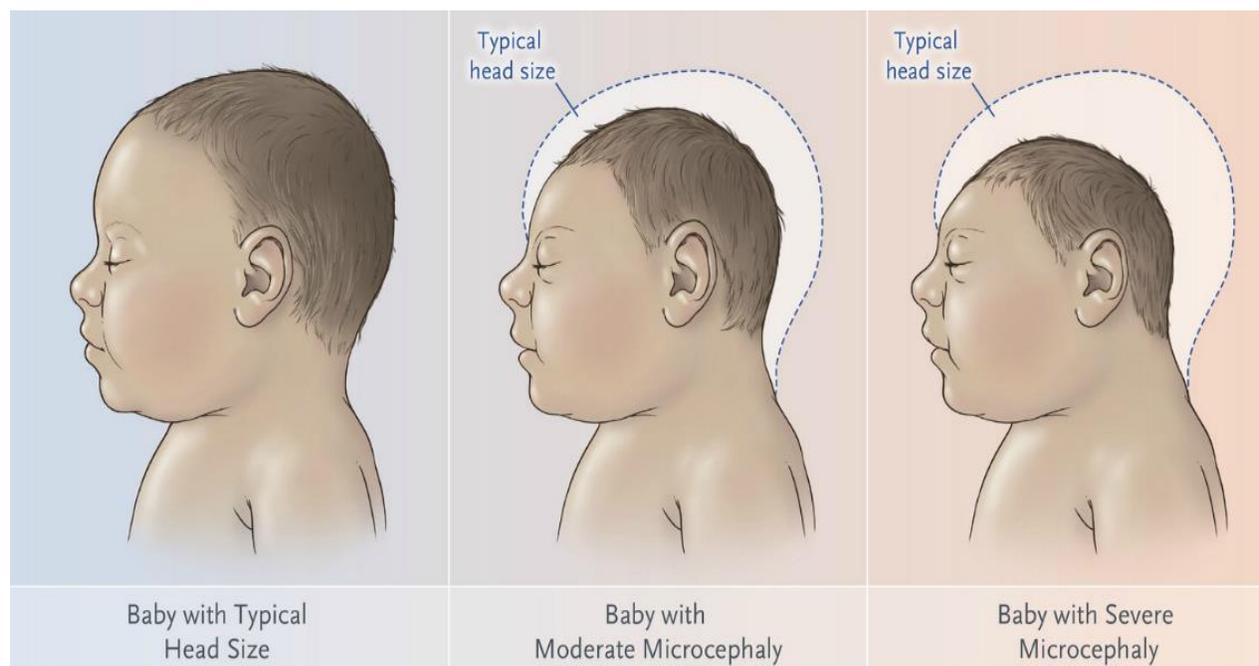


Fig. 1: Microcephaly in babies

ETIOPATHOGENESIS

ZIKV belongs to the family Flaviviridae, genus Flavivirus and is a mosquito-borne virus in the Spondweni group. Flaviviruses are arboviruses and include pathogens such as yellow fever virus (YFV), Japanese encephalitis virus (JEV), West Nile Virus (WNV), tick-borne encephalitis virus (TBEV), and the four dengue viruses (DENV1–4) in addition to ZIKV.[29]

Usually Flaviviruses are enveloped RNA viruses. They have viral genome complexed with multiple copies of the capsid protein (C) and is surrounded by an icosahedral shell composed of 180 copies of the envelope (E) glycoprotein and precursor membrane (prM) or membrane (M) proteins which are anchored in a lipid membrane. Capsid protein (C), Envelope glycoprotein (E) and Membrane protein (M) are structural proteins. When we analyze the full-length genome of ZIKV it consists of 10,794 nucleotides and encodes 3419 amino acids, in addition to the structural proteins. They constitute seven non-structural proteins (NS1, NS2a, NS2b, NS3, NS4a, NS4b, and NS5) which are involved in replication and assembly of the virus and antagonizing the host innate immune response [29,44].

NS3 and NS5 are large, multifunctional proteins. They perform several enzymatic activities involved in polyprotein processing (NS3) and RNA replication (NS3 and NS5). NS5 also have the ability to antagonize the interferon (IFN) response [39].

Mature ZIKV structure is similar to mature West Nile Virus (WNV) and Dengue virus (DENV) structures. Even though there exist certain differences in the E protein structure which are thought to be responsible for cellular tropism which contribute to various disease outcomes [44].

Phylogenetic analyses of ZIKV strains isolated have shown two distinct classes (lineages), African and Asian. The causative agent of the current ZIKV infection is the Asian lineage. It is not closely related to the African lineage but shares a common ancestor. On comparing the E protein sequences from the two lineages it has been found that the Asian lineage contains insertions in the E protein glycosylation motif which are not present in the African lineage. When the amino acid sequences of the E protein of ZIKV isolates from human, monkey and mosquito were compared, sixteen amino acid substitutions were identified, which results in subtle structural changes. These changes, may impact ZIKV virulence and host tropism [25].

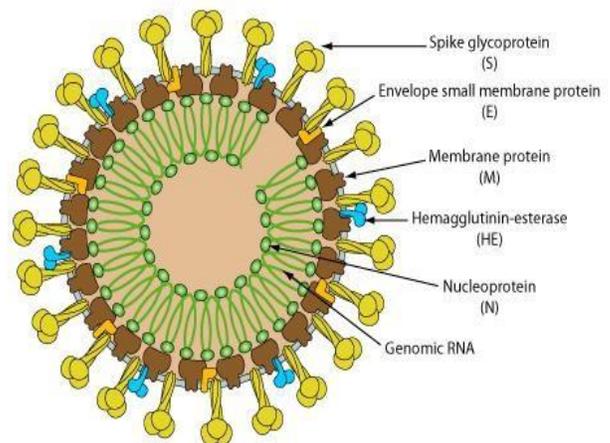


Fig. 2: Structure of Zika Virus

CLINICAL MANIFESTATION

About 80%, of individuals infected with ZIKV are asymptomatic. Pruritic maculopapular rash, low grade fever, arthritis, arthralgia, non-purulent conjunctivitis, and edema of the extremities are the most commonly found symptoms. Headache, myalgia, retro-orbital pain, low-back pain, lymphadenopathy, and vomiting are also seen in some patients. Symptoms last from several days to 1 week. Severe disease requiring hospitalization and fatalities are rare. Guillain-Barré syndrome has also been reported in patients. The disease is frequently mild but fatal cases have been reported in those with underlying medical conditions. Neurological complications associated with ZIKV infection are encephalitis, meningoencephalitis and acute myelitis. There are multiple reports of microcephaly and other birth defects following ZIKV infection during pregnancy. A causal link was suspected following the recognition of increase in the number of infants with microcephaly in Brazil and French Polynesia which were associated with outbreaks of ZIKV infection. Rasmussen *et al.* suggested sufficient evidences that had been accumulated to establish a causal relationship between ZIKV infection and microcephaly and other brain anomalies [7,11,22,39].

Evidence to support the role of ZIKV infection in causing microcephaly and other neurological abnormalities during pregnancy includes:

- Increase in microcephaly with increased ZIKV transmission
- The first trimester is the primary risk period of pregnancy according to data modelling
- ZIKV is identified in the brains of fetuses and infants (who died) with microcephaly

- (d) Human neural progenitor cell growth following in vitro ZIKV infection
- (e) in vitro infection of cytotrophoblasts and placental macrophages
- (f) Microcephaly associated with ZIKV infection in mouse and non-human primate models [34].

IMMUNITY

IFN production is a major component of the innate response. Studies show that mice deficient in various components of the IFN response have increased susceptibility to ZIKV infection. Human primary trophoblast cells, isolated from full-term placentas were found to release antiviral type III IFN λ 1 which helps to protect cells from ZIKV infection. Flaviviruses antagonize IFN signaling through multiple mechanisms. In-vitro studies show that, for proteasomal degradation, ZIKV NS5 binds to and targets the human IFN-regulated transcriptional activator STAT2. Type I and Type II IFN use different cell-surface receptors which signal through the Jak-STAT pathway, including STAT2. Thus there is possibility for ZIKV to evade Type III IFN signaling by degrading STAT2 mediated by NS5 even though they are produced by villous trophoblasts [1,27,41,48].

In humoral immunity, neutralizing antibodies have a major role in the protection against ZIKV infection. Multiple groups have identified antibodies from Dengue (DENV) patients, which cross-react, and in some cases, neutralize ZIKV. There is also concern for antibody dependent enhancement (ADE), an immunological phenomenon in which non-neutralizing or weakly neutralizing antibodies facilitate viral entrance into Fc-receptor bearing cells such as monocytes and macrophages [7,49].

Coming towards cell mediated immunity T cell responses contribute to protection and/or disease enhancement. ZIKV-specific cytokine production by CD4+ and CD8+ T cells was identified in ZIKV-infected people [33].

TRANSMISSION

- Zika virus is transmitted to people primarily through the bite of an infected *Aedes* species mosquito (*Ae. aegypti* and *Ae. albopictus*).
- Zika virus can be passed through sex from a person who has Zika to his or her sex partners.
- A pregnant woman can pass Zika virus to her fetus during pregnancy. Zika infection during pregnancy can cause serious birth defects. Zika is a cause of microcephaly and other severe fetal brain defects.

- Zika virus may be spread through blood transfusion.
- Anyone who lives in or travels to an area with risk of Zika and has not already been infected with Zika virus can get it from mosquito bites [15].



Fig.3: *Ae.aegypti* mosquito

DIAGNOSIS

Zika virus IgM antibodies can persist beyond 12 weeks after infection. We cannot reliably differentiate Zika infection occurring before pregnancy from that occurring during pregnancy from positive test results for Zika IgM.

Molecular Test for Zika Virus

Zika virus RNA can sometimes be detected early in the course of illness, for symptomatic persons with Zika virus infection. On paired serum and urine specimens RNA NAT (nucleic acid testing) is performed. NAT should be performed concurrently with IgM serology for symptomatic pregnant women with possible exposure to Zika virus. NAT is recommended three times during pregnancy for asymptomatic pregnant women with ongoing possible exposure to Zika virus.

Triplex Real-time RT-PCR Assay

For the differentiation of RNA from Zika virus and for the qualitative detection of Zika virus RNA in urine and amniotic fluid the Triplex Real-time RT-PCR Assay (TriplexrRT-PCR) is used. The assay is usually performed in specimens collected from individuals meeting CDC Zika virus clinical criteria and/or CDC Zika virus epidemiological criteria.

Serologic Test for Zika Virus

In most cases Zika virus-specific IgM and neutralizing antibodies develop toward the end of the first week of illness. IgM levels are variable, but generally are IgM is found positive from almost the day 4 after onset of symptoms and it continues up to 12 weeks after the onset of symptoms sometimes may persist longer. Thus the test for IgM is performed concurrently with NAT.

Zika MAC-ELISA

For the qualitative detection of Zika virus IgM antibodies in serum or cerebrospinal fluid, the Zika IgM Antibody Capture Enzyme-Linked Immunosorbent Assay (Zika MAC-ELISA) is used. But sometimes the results may be difficult to interpret due to the cross-reaction with other flaviviruses and possible nonspecific reactivity [15].

Nucleic acid amplification test (NAAT)

ZIKV RNA detection in the serum, whole blood, urine, and cerebrospinal fluid (CSF) are possible through NAAT. It is more specific than serological tests but have limited by the short duration of RNAemia and RNAuria. In general, ZIKV RNA can be detected from serum for approximately seven days and in urine for approximately 15–20 days of post-infection. Different NAAT have different sensitivities. Corman *et al.*, in a comparison of seven published and two new RT-PCR assays, found that due to lack of sensitivity or difficulty in obtaining necessary reagents some published RT-PCR assays may be of limited value for diagnostics in the current outbreak. Currently, 12 molecular assays are available through EUA [47].

TREATMENT

There are no specific drugs for the treatment of Zika virus.

Vaccines against Zika virus are under clinical trials. They include:

- **Inactivated Whole Organism (with or without Adjuvant)**

Inactivated vaccines against flaviviruses including JEV (Japanese Encephalitis virus) and TBEV (tick-borne encephalitis virus) have been used successfully providing support for this method. Benefits of an inactivated whole organism vaccine include multiple antigenic targets and non-replicating virus, which may improve safety. The Phase I trials of anti-ZIKV PIV vaccine candidates are under studies [32].

- **DNA vaccines**

The plasmid is injected and DNA is allowed to be taken up by antigen presenting cells, which then express it as plasmid-encoded genes and generate the target antigen. DNA vaccine against WNV (West Nile Virus) was previously tested in humans and demonstrated excellent safety and immunogenicity [28,17].

- **RNA vaccines**

RNA vaccines contain an open reading frame encoding the antigen of interest which is then translated by the host cellular machinery. A single immunization of a different ZIKV prM/E encoding mRNA lipid nanoparticle vaccine was done in rhesus macaque model and it resulted in neutralizing antibody titres that were fifty times greater than those induced by a single immunization of a DNA vaccine and more than twice as high as those induced by two immunizations of a DNA vaccine. Since there is no potential for genome integration, RNA-based vaccines may be safe than DNA vaccines [3,40].

- **Recombinant Viral Vector**

In rhesus monkeys, a rhesus adenovirus serotype 52 (RhAd52) vector-based vaccine elicited ZIKV-specific neutralizing antibodies following a single immunization which demonstrated a substantial breadth of antibody responses against linear ZIKV E protein epitopes (peptide microarray assays) [23].

- Additional vaccine candidate platforms in pre-clinical studies include live-attenuated vaccines, recombinant subunit vaccines, peptide vaccines, and ZIKV exosome vaccines [32].

Table 1: Summary of anti-ZIKA virus (ZIKV) vaccine candidates currently in clinical trials

Type of vaccine	Developers/collaborators	Candidate vaccine name (if available)	Stage of development	Clinical trial registration number
Inactivated whole organism	WRAIR/BIDMC/Harvard/NIAID/Sanofi Pasteur		Clinical (Phase I)	NCT02963909NCT02952833NCT02937233
DNA	GenOne Life Science, Inc/Inovio Pharmaceuticals	GLS-5700	Clinical (Phase I)	NCT02809443NCT02887482
DNA	VRC/NIAID	VRCZIKV DNA	Clinical (Phase I)	NCT02840487NCT02996461
Synthetic peptide	NIAID	AGS-v	Clinical (Phase I)	NCT03055000
Measles- vectored	Themis Bioscience	MV-ZIKA	Clinical (Phase I)	NCT02996890
mRNA	Valera(Moderna)	Mrna-1325	Clinical (Phase I)	NCT03014089

WRAIR:Walter Reed Army Institute of Research , BIDMC:Beth Israel Deaconess Medical Center, NIAID:National Institute of Allergy and Infectious Disease , VRC:Vaccine Research Center

No drugs are currently approved for the treatment of ZIKV-infection. The primary aim of drug development is to reduce viral load, symptoms, and to protect the unborn fetus from neurological disorders. "Re-purposing" existing compounds for the treatment of ZIKV are under studies. Multiple compounds, including ribavirin and polymerase inhibitor 7-deaza-20-C-methyladenosine (7DMA) are tested by Zmurko et al., to determine potential anti-ZIKV drugs. Of the compounds tested, 7DMA inhibited ZIKV replication in vitro, and, also reduced viremia when administered for 10 consecutive days (beginning 1 h prior to infection). It also had shown to delay time taken for disease progression in ZIKV-infected AG129 mice. But one of the main challenges is that the initiation of treatment prior to infection is impractical in non-research settings. Various other techniques such as rapid, high-throughput screening of drug/compound libraries were also utilized in order to identify compounds with in vitro anti-ZIKV activity. On analysing 727 compounds for anti-ZIKV activity using this technique, ZIKV had shown sensitivity to pyrimidine synthesis inhibitors (e.g., brequinar). Moreover, Barrows et al. screened 774 FDA-approved drugs for anti-ZIKV activity. He found that over 20 compounds, including mycophenolate mofetil, daptomycin and sertraline, are able to reduce viral infection in vitro. Then ~6000 compounds, which include approved drugs, clinical trial drug candidates, and pharmacologically active compounds, were tested to determine whether they are able to inhibit ZIKV infection or suppress infection-induced caspase-3 activity in neuronal cells. Thus it was found that emricasan, a pan-caspase

inhibitor, was the most potent anti-cell-death compound. Furthermore it has shown neuroprotective activity for human neuronal progenitor cells but they were not able to suppress ZIKV replication. Several anti-malarial compounds were also studied using a cell-based cytotoxicity assay. It had been demonstrated that they are having anti-DENV2 and anti-ZIKV properties. An FDA-approved hepatitis C virus (HCV) anti-viral, sofosbuvir was studied by Bullard-Feibelman et al., and he found that the drug was able to inhibit ZIKV replication and infection in tissue culture as well as protected mice from ZIKV-induced death [15,51].

All at a glance there are multiple challenges to be faced for the development of anti ZIKV agents. Though the clinical infection with ZIKV is typically mild, pregnant women and those at increased risk for neurological complications are the primary populations for whom treatment would be indicated. For the development of therapeutics to be used during pregnancy, there are multiple ethical considerations. In general, from clinical trials of new investigational compounds, pregnant women are excluded. But the agent should be of low risk to the mother, foetus, effective in preventing adverse effects on foetus, and practical for use in resource-limited settings [5,10].

Currently used strategies of treatment include:-

- Treat the symptoms.
- Get plenty of rest.
- Drink fluids to prevent dehydration.

- In order to reduce fever and pain take medicine such as acetaminophen.
- Do not take aspirin and other non-steroidal anti-inflammatory drugs (NSAIDS) until dengue can be ruled out to reduce the risk of bleeding.
- Talk to your healthcare provider before taking additional medication for some other medical conditions [15].

CONCLUSION:

Zika virus is a mosquito-borne flavivirus transmitted primarily by *Aedes aegypti* mosquitoes. It is a rapidly emerging virus with a complex clinical picture. Despite mild clinical symptoms in the mother, ZIKV infection during pregnancy produce harmful effects in fetus and is associated with fetal death, growth restriction, and a spectrum of central nervous system abnormalities most commonly microcephaly. Microcephaly is a condition in which baby's brain has not developed properly during pregnancy or has stopped growing after birth, which results in a smaller head size. Today, much progress has been made in understanding the epidemiology and pathogenesis of this infection. The vaccines to arrest the spread of ZIKV are under clinical trials.

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REFERENCES:

1. Abbink, P.; Larocca, R.A.; De La Barrera, R.A.; Bricault, C.A.; Moseley, E.T.; Boyd, M.; Kirilova, M.; Li, Z.; Ng'ang'a, D.; Nanayakkara, O.; et al. Protective efficacy of multiple vaccine platforms against Zikavirus challenge in rhesus monkeys. *Science* **2016**; 353: 1129–1132.
2. Adams Waldorf, K.M.; Stencel-Baerenwald, J.E.; Kapur, R.P.; Studholme, C.; Boldenow, E.; Vornhagen, J.; Baldessari, A.; Dighe, M.K.; Thiel, J.; Merillat, S.; et al. Fetal brain lesions after subcutaneous inoculation of Zika virus in a pregnant nonhuman primate. *Nat. Med.* **2016**; 22: 1256–1259.
3. Adcock, R.S.; Chu, Y.K.; Golden, J.E.; Chung, D.H. Evaluation of anti-Zika virus activities of broad-spectrum antivirals and NIH clinical collection compounds using a cell-based, high-throughput screen assay. *Antivir. Res.* **2017**; 138: 47–56.
4. Azevedo, R.S.; Araujo, M.T.; Martins Filho, A.J.; Oliveira, C.S.; Nunes, B.T.; Cruz, A.C.; Nascimento, A.G.; Medeiros, R.C.; Caldas, C.A.; Araujo, F.C.; et al. Zika virus epidemic in Brazil. I. Fatal disease in adults: Clinical and laboratorial aspects. *J. Clin. Virol.* **2016**; 85: 56–64.
5. Balasubramanian, A.; Teramoto, T.; Kulkarni, A.A.; Bhattacharjee, A.K.; Padmanabhan, R. Antiviral activities of selected antimalarials against dengue virus type 2 and Zika virus. *Antivir. Res.* **2017**; 137: 141–150.
6. Bayer, A.; Lennemann, N.J.; Ouyang, Y.; Bramley, J.C.; Morosky, S.; Marques, E.T., Jr.; Cherry, S.; Sadovsky, Y.; Coyne, C.B. Type III Interferons Produced by Human Placental Trophoblasts Confer Protection against Zika Virus Infection. *Cell Host Microbe* **2016**; 19: 705–712.
7. Barba-Spaeth, G.; Dejnirattisai, W.; Rouvinski, A.; Vaney, M.C.; Medits, I.; Sharma, A.; Simon-Loriere, E.; Sakuntabhai, A.; Cao Lormeau, V.M.; Haouz, A.; et al. Structural basis of potent Zika-dengue virus antibody cross-neutralization. *Nature* **2016**; 536: 48–53.
8. Boeuf, P.; Drummer, H.E.; Richards, J.S.; Scoullar, M.J.; Beeson, J.G. The global threat of Zika virus to pregnancy: Epidemiology, clinical perspectives, mechanisms, and impact. *BMC Med.* **2016**; 14: 112.
9. Brasil P, Pereira JP Jr, Raja Gabaglia C, et al. Zika virus infection in pregnant women in Rio de Janeiro — preliminary report. *N Engl J Med.* DOI: 10.1056/NEJMoa1602412
10. Bullard-Feibelman, K.M.; Govero, J.; Zhu, Z.; Salazar, V.; Veselinovic, M.; Diamond, M.S.; Geiss,

- B.J.The FDA-approved drug sofosbuvir inhibits Zika virus infection. *Antivir. Res.* **2017**; 137: 134–140.
- 11.Campos GS, Bandeira AC, Sardi SI. Zika virus outbreak, Bahia, Brazil. *Emerg Infect Dis* 2015;21:1885–6.
<http://dx.doi.org/10.3201/eid2110.150847>.
- 12.Calvet, G.A.; Santos, F.B.; Sequeira, P.C. Zika virus infection: Epidemiology, clinical manifestations and diagnosis. *Curr. Opin. Infect. Dis.* **2016**; 29: 459–466.
- 13.Carteaux, G.; Maquart, M.; Bedet, A.; Contou, D.; Brugieres, P.; Fourati, S.; Cleret de Langavant, L.; deBroucker, T.; Brun-Buisson, C.; Leparco-Goffart, I.; et al. Zika Virus Associated with Meningoencephalitis. *N. Engl. J. Med.* **201**; 374: 1595–1596.
- 14.Carvalho, M.D.; Miranda-Filho, D.B.; van der Linden, V.; Sobral, P.F.; Ramos, R.C.; Rocha, M.A.; Cordeiro, M.T.; de Alencar, S.P.; Nunes, M.L. Sleep EEG patterns in infants with congenital Zika virus syndrome. *Clin. Neurophysiol.* **2016**; 128: 204–214.
- 15.Centers for Disease Control and Prevention National Center for Emerging and Zoonotic Infectious Diseases (NCEZID) Division of Vector-Borne Diseases (DVBD)
- 16.D’Ortenzio, E.; Matheron, S.; Yazdanpanah, Y.; de Lamballerie, X.; Hubert, B.; Piorowski, G.; Maquart, M.; Descamps, D.; Damond, F.; Leparco-Goffart, I. Evidence of Sexual Transmission of Zika Virus. *N. Engl. J. Med.* **2016**; 374: 2195–2198.
- 17.Dowd, K.A.; Ko, S.Y.; Morabito, K.M.; Yang, E.S.; Pelc, R.S.; DeMaso, C.R.; Castilho, L.R.; Abbink, P.; Boyd, M.; Nityanandam, R.; et al. Rapid development of a DNA vaccine for Zika virus. *Science* **2016**; 354: 237–240.
- 18.Foy, B.D.; Kobylinski, K.C.; Chilson Foy, J.L.; Blitvich, B.J.; Travassos da Rosa, A.; Haddow, A.D.; Lanciotti, R.S.; Tesh, R.B. Probable non-vector-borne transmission of Zika virus, Colorado, USA. *Emerg. Infect. Dis.* **2011**; 17: 880–882.
- 19.Frank, C.; Cadar, D.; Schlaphof, A.; Neddersen, N.; Gunther, S.; Schmidt-Chanasit, J.; Tappe, D. Sexual transmission of Zika virus in Germany, April 2016. *Euro Surveill.* **2016**; 21.
- 20.Frank, C.; Faber, M.; Stark, K. Causal or not: Applying the Bradford Hill aspects of evidence to the association between Zika virus and microcephaly. *EMBO Mol. Med.* **2016**; 8: 305–307.
- 21.Freour, T.; Mirallie, S.; Hubert, B.; Spingart, C.; Barriere, P.; Maquart, M.; Leparco-Goffart, I. Sexual transmission of Zika virus in an entirely asymptomatic couple returning from a Zika epidemic area, France, April 2016. *Euro Surveill.* **2016**; 21.
- 22.Grossi-Soyster, E.N.; LaBeaud, A.D. Clinical aspects of Zika virus. *Curr. Opin. Pediatr.* **2016**.
- 23.Halstead, S.B. Antibodies determine virulence in dengue. *Ann. N. Y. Acad. Sci.* **2009**, 1171, E48–E56.
- 24.Honein MA, Rasmussen SA, Jamieson DJ, Petersen LR. Zika virus and birth defects — reviewing the evidence for causality. *N Engl J Med* 2016;374:1981-1987
- 25.Kindhauser, M.K.; Allen, T.; Frank, V.; Santhana, R.S.; Dye, C. Zika: The origin and spread of a mosquito-borne virus. *Bull. World Health Organ.* **2016**; 94: 675–686.
- 26.Kleber de Oliveira, W.; Cortez-Escalante, J.; de Oliveira, W.T.; do Carmo, G.M.; Henriques, C.M.; Coelho, G.E.; Araujo de Franca, G.V. Increase in Reported Prevalence of Microcephaly in Infants Born to Women Living in Areas with Confirmed Zika Virus Transmission During the First Trimester of Pregnancy—Brazil, 2015. *MMWR Morb. Mortal. Wkly. Rep.* **2016**; 65: 242–247.
- 27.Kotenko, S.V.; Gallagher, G.; Baurin, V.V.; Lewis-Antes, A.; Shen, M.; Shah, N.K.; Langer, J.A.; Sheikh, F.; Dickensheets, H.; Donnelly, R.P. IFN- λ mediate antiviral protection through a distinct class II cytokine receptor complex. *Nat. Immunol.* **2003**; 4: 69–77.
- 28.Kramps, T.; Elbers, K. Introduction to RNA Vaccines. *Methods Mol. Biol.* **2017**; 1499: 1–11.
- 29.Kuno, G.; Chang, G.J. Full-length sequencing and genomic characterization of Bagaza, Kedougou, and Zika viruses. *Arch. Virol.* **2007**; 152: 687–696.
- 30.Landry, M.L.; St George, K. Laboratory Diagnosis of Zika Virus Infection. *Arch. Pathol. Lab. Med.* **2017**; 141:60–67.
- 31.Lazear, H.M.; Govero, J.; Smith, A.M.; Platt, D.J.; Fernandez, E.; Miner, J.J.; Diamond, M.S. A Mouse Model of Zika Virus Pathogenesis. *Cell Host Microbe* **2016**; 19: 720–730.
- 32.Ledgerwood, J.E.; Pierson, T.C.; Hubka, S.A.; Desai, N.; Rucker, S.; Gordon, I.J.; Enama, M.E.; Nelson, S.; Nason, M.; Gu, W.; et al. A West Nile virus DNA vaccine utilizing a modified promoter induces neutralizing antibody in younger and older healthy adults in a phase I clinical trial. *J. Infect. Dis.* **2011**; 203: 1396–1404.
- 33.Lehrer, A.T.; Holbrook, M.R. Tick-borne Encephalitis Vaccines. *J. Bioterror. Biodef.* 2011, 3.
- 34.Martin, J.E.; Pierson, T.C.; Hubka, S.; Rucker, S.; Gordon, I.J.; Enama, M.E.; Andrews, C.A.; Xu, Q.; Davis, B.S.; Nason, M.; et al. A West Nile virus DNA vaccine induces neutralizing antibody in healthy adults during a phase I clinical trial. *J. Infect. Dis.* **2007**; 196: 1732–1740.
- 35.Martines, R.B.; Bhatnagar, J.; de Oliveira Ramos, A.M.; Davi, H.P.; Iglezias, S.D.; Kanamura, C.T.; Keating, M.K.; Hale, G.; Silva-Flannery, L.; Muehlenbachs, A.; et al. Pathology of congenital

- Zika syndrome in Brazil: A case series. *Lancet* **2016**; 388: 898–904.
36. Mecharles, S.; Herrmann, C.; Poullain, P.; Tran, T.H.; Deschamps, N.; Mathon, G.; Landais, A.; Breurec, S.; Lannuzel, A. Acute myelitis due to Zika virus infection. *Lancet* **2016**; 387: 1481.
37. Mlakar, J.; Korva, M.; Tul, N.; Popovic, M.; Poljsak-Prijatelj, M.; Mraz, J.; Kolenc, M.; Resman Rus, K.; Vesnaver Vipotnik, T.; Fabjan Vodusek, V.; et al. Zika Virus Associated with Microcephaly. *N. Engl. J. Med.* **2016**; 374: 951–958.
38. Quicke, K.M.; Bowen, J.R.; Johnson, E.L.; McDonald, C.E.; Ma, H.; O’Neal, J.T.; Rajakumar, A.; Wrammert, J.; Rimawi, B.H.; Pulendran, B.; et al. Zika Virus Infects Human Placental Macrophages. *Cell Host Microbe* **2016**; 20: 83–90.
39. Ramaiah, A.; Dai, L.; Contreras, D.; Sinha, S.; Sun, R.; Arumugaswami, V. Comparative analysis of protein evolution in the genome of pre-epidemic and epidemic Zika virus. *Infect. Genet. Evol.* **2017**; 51: 74–85.
40. Salam, A.P.; Rojek, A.; Dunning, J.; Horby, P.W. Clinical Trials of Therapeutics for the Prevention of Congenital Zika Virus Disease: Challenges and Potential Solutions. *Ann. Intern. Med.* **2017**.
41. Sapparapu, G.; Fernandez, E.; Kose, N.; Bin, C.; Fox, J.M.; Bombardi, R.G.; Zhao, H.; Nelson, C.A.; Bryan, A.L.; Barnes, T.; et al. Neutralizing human antibodies prevent Zika virus replication and fetal disease in mice. *Nature* **2016**; 540: 443–447.
42. Schneider, W.M.; Chevillotte, M.D.; Rice, C.M. Interferon-stimulated genes: A complex web of host defenses. *Annu. Rev. Immunol.* **2014**; 32: 513–545.
43. Sharma, A.; Lal, S.K. Zika Virus: Transmission, Detection, Control, and Prevention. *Front. Microbiol.* **2017**; 8: 110.
44. Sirohi, D.; Chen, Z.; Sun, L.; Klose, T.; Pierson, T.C.; Rossmann, M.G.; Kuhn, R.J. The 3.8 Å resolution cryo-EM structure of Zika virus. *Science* **2016**; 352: 467–470.
45. Soares, C.N.; Brasil, P.; Carrera, R.M.; Sequeira, P.; de Filippis, A.B.; Borges, V.A.; Theophilo, F.; Ellul, M.A.; Solomon, T. Fatal encephalitis associated with Zika virus infection in an adult. *J. Clin. Virol.* **2016**; 83: 63–65.
46. Tang, H.; Hammack, C.; Ogden, S.C.; Wen, Z.; Qian, X.; Li, Y.; Yao, B.; Shin, J.; Zhang, F.; Lee, E.M.; et al. Zika Virus Infects Human Cortical Neural Progenitors and Attenuates Their Growth. *Cell Stem Cell* **2016**; 18: 587–590.
47. US Food & Drug Administration Emergency Use Authorizations. Available online <https://www.fda.gov/MedicalDevices/Safety/EmergencySituations/ucml61496.htm#zika> (accessed on 7 March 2017).
48. Versteeg, G.A.; Garcia-Sastre, A. Viral tricks to grid-lock the type I interferon system. *Curr. Opin. Microbiol.* **2010**; 13: 508–516.
49. WHO Vaccine Pipeline Tracker Zika. Available online: http://www.who.int/immunization/research/vaccine_pipeline_tracker_spreadsheet/en/ (accessed on 14 March 2017).
50. WHO Situation Report 10 March 2017. Available online: <http://apps.who.int/iris/bitstream/10665/254714/1/zik-asitrep10Mar17-eng.pdf?ua=1> (accessed on 2 April 2017).
51. Zmurko, J.; Marques, R.E.; Schols, D.; Verbeken, E.; Kaptein, S.J.; Neyts, J. The Viral Polymerase Inhibitor 7-Deaza-20-C-Methyladenosine Is a Potent Inhibitor of In Vitro Zika Virus Replication and Delays Disease Progression in a Robust Mouse Infection Model. *PLoS Negl. Trop. Dis.* **2016**; 10, e0004695.